



Review Article



Green Synthesis of Silver Nanoparticle from Different Plants– A Review

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Received: 15.02.2016 | Revised: 26.02.2016 | Accepted: 12.03.2016

ABSTRACT

Nanotechnology is advance impetus due to its capacity to transform metals into nano particles. Among the metal nano particles, silver nano particles play a vital role in antimicrobial activity, a very important application known from ancient times. A systemic characterization of synthesized silver nano particles was performed using UV, SEM, TEM, XRD and EDX. In this communication, we report a review on the green method for synthesis of nano particles.

Key words: Nanotechnology, Nanoparticle, Metal nanoparticle, Green chemistry, silver nanoparticle, antimicrobial activity.

INTRODUCTION

Nanotechnology is rapidly growing filed throughout the world and expected to grow due to wide a number of applications being conducted. Nanotechnology and nano biotechnology are the two emerging filed to improve the product being manufacture and marketed¹. Nanotechnology is defined as manipulation of matter on atomic and molecular scales. Nanotechnology is classified into two forms, Wet and Dry nanotechnology. Wet nanotechnology deals with living biosynthesis Dry nanotechnology deals with man-made objects². Nano biotechnology is a field that has emerged as an interaction between the two advanced areas such as biotechnology and nanotechnology for developing new bio synthetic devices and eco friendly technology for the synthesis of nano materials in nano scale³. Nano particle are the fundamental building blocks of nanotechnology. Particles with size up to 100nm are usually referred as

nano particles. Nano particles have greater applications. The greater attention towards the nano particles is that it is not only due to various application and also by its way of synthesis in different filed⁴. Nanoparticles synthesis can be achieved through various approaches including solution reduction, chemical, photochemical, reverse micelles thermal decomposition, radiation assisted, electrochemical, microwave assisted method and recently via biological routes⁵⁻⁹. Historically the most effectively studied nanoparticles are mostly derived from noble metals in particular gold, platinum, copper, zinc, titanium, selenium, magnesium, silver and alginate¹⁰. Metal nano particle have tremendous application in the field of medicine, defense, drug synthesis, catalysis, diagnostics, electronics, biological probes, chemical and bio chemical sensing optics. To date among the above metal nanoparticles, silver nano particle plays an important role in nanotechnology¹¹.

Cite this article: Asha, A., Sivarajani, T., Thirunavukkarasu, P. and Asha, S., Green Synthesis of Silver Nanoparticle from Different Plants– A Review, *Int. J. Pure App. Biosci.* **4**(2): 118-124 (2016). doi: <http://dx.doi.org/10.18782/2320-7051.2221>

Silver nanoparticle, a noble metal nano particle focus it to have a wide ranging application in area of catalysis, antimicrobial, chemical sensing, photochemistry, optoelectronics, electrical conducting and bio material production¹². Silver nanoparticles can be synthesized by using various approaches including chemical, physical and biological method. In recent time biosynthetic method has received more attention than chemical and physical methods. Among this bio material used biological synthesis includes usage of plant (or) microbes (bacteria, fungi, algae). Synthesis of nanoparticles using plant mediated synthesis as more importance than using microbes due to adverse microbial culture maintenance, time

consume and cost effective. For synthesis of nanoparticle, plant extracts utilization as gained tremendous influence in large scale bio synthesis of nanotechnology. The green chemistry synthesis has exhibited a clean, non toxic, non hazardous, cost effective, environmental friendly and therapeutics of extensive use to humans since chemical methods are extremely expensive and consumed toxic chemical which may inform potent environmental and biological risk factors¹³. There are so many reports related to the synthesis of bioactive silver nano particles using several plants extract has been already documented in various approaches is given in the following table.

Table 1:

| S.No | Name of the medicinal Species | Part's | Extract | N.P | Particle size (nm) | Particle shape | Methods | Name of the Microorganism | Zone diameter (mm) |
|------|---|--------|---------|-----|--------------------|----------------|---------------------|-------------------------------------|--------------------|
| 1. | <i>Acacia nilotica</i> ¹⁴ | Leaf | Aqueous | Ag | 30-150 | - | Well diffusion | <i>Pseudomonas</i> | 26 |
| | | | | | | | | <i>Escherichia coli</i> | 20 |
| | | | | | | | | <i>Basillus sp</i> | 23 |
| | | | | | | | | <i>Proteus sp</i> | 13 |
| | | | | | | | | <i>Escherichia coli</i> | 22 |
| 2. | <i>Annona reticulata</i> ¹⁵ | Leaf | Aqueous | Ag | 20-30 | Spherical | Agar well diffusion | <i>Staphylococcus aureus</i> | 20 |
| | | | | | | | | <i>Escherichia coli</i> | 22 |
| 3. | <i>Arbutus unedo</i> ¹⁶ | Leaf | Aqueous | Ag | 9-15 | Spherical | Well diffusion | <i>Escherichia coli</i> | 7 |
| | | | | | | | | <i>Pseudomonas putida</i> | 5 |
| | | | | | | | | <i>Klebesiella pneumoniae</i> | 5 |
| | | | | | | | | <i>Bacillus subtilis</i> | 3 |
| | | | | | | | | <i>Staphylococcus Aureus</i> | 8 |
| | | | | | | | | <i>Escherichia coli</i> | 16 |
| 4. | <i>Argyeria nervosa</i> ¹⁷ | Seed | Aqueous | Ag | 20-50 | Spherical | Agar Well diffusion | <i>Staphylococcus aureus</i> | 14 |
| | | | | | | | | <i>Bacillus subtilis</i> | 17 |
| | | | | | | | | <i>Pichla pastoris</i> | 12 |
| | | | | | | | | <i>Aspergillus niger</i> | 17 |
| | | | | | | | | <i>Echcherichia coli</i> | 16 to 20 |
| 5. | <i>Arnebia nobilis</i> ¹⁸ | Root | Aqueous | Ag | 40-70 | Spherical | Well diffusion | <i>Staphlococcus aureus</i> | 15 to 19 |
| | | | | | | | | <i>Basillus cereus</i> | 9 |
| 6. | <i>Artocarpus heterophyllus Lam</i> ¹⁹ | Seed | Powder | Ag | 3-25 to 10.78 | Irregular | Agar diffusion | <i>Bacillus subtilis</i> | 12 |
| | | | | | | | | <i>Staphylococcus aureus</i> | 15 |
| | | | | | | | | <i>Pseudomonas aeruginosa</i> | 6 |
| | | | | | | | | <i>Bacillus subtilis</i> | 29 |
| 7. | <i>Banana</i> ²⁰ | Peel | Aqueous | Ag | 23.7 | Spherical | Well diffusion | <i>Staphylococcus aureus</i> | 36 |
| | | | | | | | | <i>Pseudomons aeruginosa(ATC C)</i> | 40 |
| | | | | | | | | <i>Pseudomonas aeruginosa</i> | 38 |
| | | | | | | | | <i>Escherichia coli</i> | 39 |

| | | | | | | | | | |
|-----|---|--------|---------|----|----------|----------------------|----------------------------|---------------------------------|---------------|
| 8. | <i>Ceratonia siliqua</i> ²¹ | Leaf | Aqueous | Ag | 5-40 | Spherical 1 | Disc diffusion | <i>Escherichia coli</i> | 8to12 |
| 9. | <i>Citrus sinesis</i> ²² | Peel | Aqueous | Ag | 80 | Square and rectangle | Disc diffusion | <i>Staphyillus epidermidis</i> | 12 |
| | | | | | | | | <i>Bacillus cereus</i> | 10 |
| | | | | | | | | <i>F.acuminatum</i> | 6 |
| | | | | | | | | | |
| 10. | <i>Coriander sativum</i> ²³ | Leaf | Aqueous | Ag | 8 | Spherical 1 | - | <i>Staphylococcus aureus</i> | 28 |
| | | | | | | | | <i>Klebsiella pneumonia</i> | 17 |
| | | | | | | | | <i>Escherichia coli</i> | 21 |
| | | | | | | | | <i>Pseudomonas aeruginosa</i> | 27 |
| | | | | | | | | <i>Escherichia coli</i> | 9.0 |
| 11. | <i>Crataegus douglasii</i> ²⁴ | Fruit | Aqueous | Ag | 29.28 | Spherical 1 | Disc diffusion | <i>Staphylococcus aureus</i> | 13.0 |
| 12. | <i>Eucalyptus chapmanian a</i> ²⁵ | Leaves | Aqueous | Ag | - | Crystal in nature | Well diffusion | <i>Staphylococcus aureus</i> | 27 |
| | | | | | | | | <i>Staphylococcus pneumonia</i> | 25 |
| | | | | | | | | <i>Klebsiella pneumonia</i> | 23 |
| | | | | | | | | <i>Escherichia coli</i> | 23 |
| | | | | | | | | <i>P.vulgaris</i> | 23 |
| | | | | | | | | <i>Candida albicans</i> | 25 |
| | | | | | | | | | |
| 13. | <i>Manikara zapota</i> ²⁶ | Seed | Aqueous | Ag | 5-35 | Circular | Agar diffusion | <i>Candida albicans</i> | 24 |
| | | | | | | | | <i>Candida tropicalis</i> | 15 |
| | | | | | | | | <i>Candida krusei</i> | 14 |
| | | | | | | | | <i>Candida gulliemondi</i> | 17 |
| | | | | | | | | <i>Candida lusitaniae</i> | 13 |
| | | | | | | | | | |
| 14. | <i>Mimusops elengi</i> ²⁷ | Leaf | Aqueous | Ag | 55-83 | Spherical 1 | Kirby Bauer Disc diffusion | <i>Klebsiella Pneumonia</i> | 8to18 |
| | | | | | | | | <i>Staphylococcus aureus</i> | 6.75 to 8 |
| | | | | | | | | <i>Micrococcus lutes</i> | 7 to 11 |
| 15. | <i>Morianda citrifolia</i> ²⁸ | Leaf | Aqueous | Ag | 22-193.5 | Spherical 1 | Agar well diffusion | <i>Candida tropicalis</i> | 28.0-57 |
| | | | | | | | | <i>Candida albicans</i> | 24.0-1.52 |
| 16. | <i>Ocimum sanctum</i> ²⁹ | Leaves | Aqueous | Ag | 18 | Spherical 1 | Kirby Bauer Disc diffusion | <i>Escherichia coli</i> | 11 |
| | | | | | | | | <i>Staphylococcus aureus</i> | 10 |
| 17. | <i>Parthenium hysterophorus</i> ³⁰ | Leaf | Aqueous | Ag | 30-80 | Irregular | Disc diffusion | <i>Salmonella sp</i> | 0.4 to 1.2 |
| | | | | | | | | <i>Staphylococcus sp</i> | 0.3 to 0.8 |
| | | | | | | | | <i>Pseudomonas sp</i> | 0.2 to 0.7 |
| | | | | | | | | <i>Escherichia sp</i> | 0.1 to 0.5 |
| | | | | | | | | <i>Penicillium sp</i> | 0.1 to 0.3 |
| | | | | | | | | | |
| 18. | <i>Piper longum</i> ³¹ | Fruit | Aqueous | Ag | 46 | Spherical 1 | Disc diffusion | <i>Staphylococcus aureus</i> | 1.0 to 1.4 |
| | | | | | | | | <i>Bacillus aureus</i> | 0.8 to 1.3 |
| | | | | | | | | <i>Pseudomonas</i> | 0 to 1.8 |
| | | | | | | | | <i>Bacillus subtilis</i> | 0 to 2.0 |
| 19. | <i>Phyllanthus amarus</i> ³² | Leaf | Aqueous | Ag | 29.78 | Spherical 1 | Agar well diffusion | <i>P.aeruginosa</i> | 10 to 21 |
| 20. | <i>Sesbania grandiflora</i> ³³ | Leaf | Aqueous | Ag | 10-25 | Spherical 1 | Disc diffusion | <i>Salmonella enteric</i> | 15.67 to 0.9 |
| | | | | | | | | <i>Staphylococcus</i> | 10.54 to 0.23 |

| | | | | | | | | | |
|-----|---|------------------|---------|-----------|------------------------------|---|----------------|--|----------------------------|
| 21. | <i>Tribulus terrestris</i> ³⁴ | Fruit | Aqueous | Ag | 16-28 | Spherica 1 | Kirby Bauer | <i>Staphylococcus aureus</i> | 9.75 |
| | | | | | | | | <i>Bacillus subtilis</i> | 9.25 |
| | | | | | | | | <i>Escherichia coli</i> | 10.75 |
| | | | | | | | | <i>Pseudomonas aeruginosa</i> | 9.25 |
| | | | | | | | | <i>Staphylococcus pyogenes</i> | 10 |
| | | | | | | | | Au | Ag |
| 22. | <i>Trianthema decandra L</i> ³⁵ | Isolated saponin | Aqueous | Au and Ag | Au=37.7-79.9 Ag=17.9-59.6 | Au-Spherica 1, Cubical, hexagonal Ag-Spherica 1 | Kirby Bauer | <i>Enterococcus faecalis</i> <i>Staphylococcus aureus</i> <i>Staphylococcus faecalis</i> <i>Bacillus subtilis</i> | 8.2 to 11.5 7.8 to 20.3 |
| | | | | | | | | | |
| | | | | | | | | | |
| | | | | | | | | | |
| | | | | | | | | | |
| 23. | <i>Xanthan</i> ³⁶ | Gum | - | Ag | 5-40 | Spherica 1 | Disc diffusion | <i>Staphylococcus aureus</i> | 12.3 to 12.6 |
| | | | | | | | | <i>Escherichia coli</i> | 9.7 to 10.7 |
| 24. | <i>Ficus microcarpa</i> ³⁷ | Leaf | Aqueous | Ag | 100 | - | Disc diffusion | <i>Bacillus cereus</i> | 10 |
| | | | | | | | | <i>Escherichia coli</i> | 12 |
| | | | | | | | | <i>Klebsiella pneumoniae</i> | 12 |
| 25. | <i>Lantana camara</i> ³⁸ | Leaf | Aqueous | Ag | - | Spherica 1 | Well diffusion | <i>Bacillus subtilis</i> | 11 to 20 |
| | | | | | | | | <i>Staphylococcus aureus</i> | 10 to 18 |
| | | | | | | | | <i>Pseudomonas aeruginosa</i> | 12 to 17 |
| | | | | | | | | <i>Escherichia coli</i> | 14 to 17 |
| 26. | <i>Cochlospermum Religiosum</i> ³⁹ | Leaf | Aqueous | Ag | 40-100 | Spherica 1 | Disc diffusion | <i>Klebsiella Pseudomonas</i> | 11 |
| | | | | | | | | <i>Basillus</i> | 12 |
| | | | | | | | | <i>Staphylococcus</i> | 15 |
| | | | | | | | | <i>Escherichia coli</i> | 20 |
| | | | | | | | | <i>Escherichia coli</i> | 88 |
| 27. | <i>Euphorbia hirta</i> ⁴⁰ | Leaf | Aqueous | Ag | 6-71 | Spherica 1 | Well diffusion | <i>Pseudomonas Aeroginosa</i> | 86 |
| | | | | | | | | <i>Klebsiella pneumonia</i> | 94 |

CONCLUSION

Thus, in the present review, the synthesis of silver nanoparticle is possible due to reduction of ion by compounds present in the plant extract. The synthesized silver nano particle using “green chemistry” are safe, effective, environmental friendly and biomedical importance. The ancient system of medicine has reported that silver based nanomedicine plays an important role. Silver nanoparticles have a lot of therapeutic and social values due to its antimicrobial properties since microorganisms are the major source of diseases.

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